Comparative effects of two different forms of selenium on oxidative stress biomarkers in healthy men: a randomized clinical trial

John P. Richie Jr,
Departments of Public Health Sciences, Pennsylvania State University College of Medicine, Hershey, PA

Arun Das,
Biochemistry and Molecular Biology, Pennsylvania State University College of Medicine, Hershey, PA

Ana M. Calcagnotto,
Departments of Public Health Sciences, Pennsylvania State University College of Medicine, Hershey, PA

Raghu Sinha,
Biochemistry and Molecular Biology, Pennsylvania State University College of Medicine, Hershey, PA

Wanda Neidig,
Penn State Hershey Cancer Institute Clinical Trials Office, Pennsylvania State University College of Medicine, Hershey, PA

Jiangang Liao,
Departments of Public Health Sciences, Pennsylvania State University College of Medicine, Hershey, PA

Eugene J. Lengerich,
Departments of Public Health Sciences, Pennsylvania State University College of Medicine, Hershey, PA

Arthur Berg,
Departments of Public Health Sciences, Pennsylvania State University College of Medicine, Hershey, PA

Corresponding Author: Karam El-Bayoumy, Ph.D., Department of Biochemistry and Molecular Biology, Penn State University College of Medicine, Penn State Hershey Cancer Institute, 500 University Drive, PO Box 850, Hershey, PA 17033, Phone: 717-531-1005; kelbayoumy@hmc.psu.edu.

The authors’ responsibilities were as follows: JPR, KE-B, RS, and JL conceived and designed the study with input from all authors; AD, AC, RS, WN, AC, WN, SN, and AMC, conducted the research; JPR, RS, JL, AB, KE-B, and TJH analyzed the data and performed statistical analyses; JPR, RS and KE-B wrote the paper with help from all authors; all authors read and approved the final manuscript.

Conflict of Interest: Selenium yeast supplements (SeleneExcell®) and partial research support for selenium speciation analyses were provided by Cypress Systems, Inc. (Madera, CA). Cypress Systems, Inc. had no role in the study design and conduct, collection, management, analysis, and interpretation of the data, or writing of the report. They also had no access to the database. None of the other authors have any conflicts of interest to disclose.
Abstract

Epidemiological and laboratory studies indicate that dietary selenium protects against prostate cancer. Results from clinical trials suggest that selenium-enriched yeast (SY) but not selenomethionine (SeMet) may be effective at reducing prostate cancer risk. Our objectives were to directly compare for the first time the effects of SeMet and SY on prostate cancer relevant biomarkers in men. We performed a randomized double blind, placebo-controlled trial of SY (200 or 285 µg/day) and SeMet (200 µg/day) administered for 9 months in 69 healthy men. Primary endpoints included blood levels of selenium-containing compounds and oxidative stress biomarkers (urine 8-hydroxy-2'-deoxyguanosine [8-OHdG] and 8-iso-prostaglandin-F$_2$α [8-iso-PGF$_2$α] and blood glutathione [GSH]). Secondary endpoints included plasma glucose and PSA levels. Compliance was high in all groups (>95%). Plasma selenium levels were increased 93%, 54%, and 86% after 9 months in SeMet and low and high dose SY groups, respectively, and returned to baseline levels after a 3 month washout (P<0.05). Levels of 8-OHdG and 8-iso-PGF$_2$α were decreased 34% and 28%, respectively, after 9 months in the high dose SY group (P<0.05). These decreases were greatest in individuals with low baseline plasma levels of selenium (<127 ng/ml). No changes in serum PSA or blood glucose and GSH were observed. Overall, we showed for the first time, reductions in biomarkers of oxidative stress following supplementation with SY but not SeMet in healthy men. These findings suggest that selenium-containing compounds other than SeMet may account for the decrease in oxidative stress.

Keywords

selenium-enriched yeast; selenomethionine; prostate cancer; oxidative stress
Introduction

Prostate cancer (PC) is the second leading cause of cancer-related deaths in men (1). Diet-derived agents including selenium have been shown to have chemopreventive potential against PC (2, 3). Epidemiological and laboratory investigations have shown that dietary selenium is protective against the development of cancer at many sites including the prostate (3–7). In the Nutritional Prevention of Cancer (NPC) study (8), supplementation with selenium-enriched yeast (SY) in men was associated with an ~50% reduction in cancer morbidity and mortality, including a 63% decrease in PC incidence, with subjects having low baseline plasma selenium levels (<122 ng/ml) showing the greatest benefit (9). However, in the Selenium and Vitamin E Cancer Prevention Trial (SELECT), which was designed to test the protective effects of selenomethionine (SeMet) and vitamin E individually and in combination against PC (10), no protection was observed, supporting the notion that SeMet is a form of selenium which is not highly active against PC (10). This is in line with laboratory studies which have consistently demonstrated that while multiple organic forms of selenium have anticancer activity, SeMet alone was relatively inactive (11). These results suggest that while SeMet represents a major form of selenium in SY, it is not likely the form responsible for the chemopreventive properties of SY.

While results from previous studies support the chemoprotective efficacy of SY (8, 9) but not SeMet (10), no direct comparison between SY and SeMet supplementation in men has been reported. Direct comparisons in laboratory animals have revealed differing results. In the rat, McCormick et al (12) reported the lack of protective effect of both SeMet and SY against the development of PC. In dogs, no differences were observed in tissue selenium levels or in the levels of several biomarkers of prostate epithelial DNA damage, proliferation or apoptosis between SY and SeMet supplemented groups (13). In contrast to the above mentioned studies, supplementation with different forms of selenium (selenite, SeMet and SY) resulted in clear differences in gene expression profiles in mice; SY was the only form associated with a pattern of decreased DNA damage (14).

In order to directly compare the effects of SY and SeMet for the first time in humans, we conducted a double blind, placebo-controlled, randomized trial in healthy adult men. Our primary objectives were to determine and compare the effects of these different forms of selenium on PC relevant biomarkers including blood selenium levels and blood and urinary biomarkers of oxidative stress. A secondary objective in this trial was to determine the impact of SY and SeMet on PSA and glucose levels.

Methods

Trial design

The study (ClinicalTrials.gov: NCT01112449) was conducted with approval from the Institutional Review Boards of the Penn State University (PSU) College of Medicine and Robert Wood Johnson Medical School. Recruitment, interviewing and sample collections were performed at three sites: Penn State Hershey Cancer Institute, Hershey, PA, PSU Clinical Research Center, State College, PA and the Cancer Institute of New Jersey, New Brunswick, NJ. Subjects were recruited from local surrounding areas using fliers, newspaper
and radio advertisements, online announcements and word of mouth. Potentially eligible subjects as assessed by telephone prescreening, were interviewed in the clinic after providing informed consent. Each subject was screened for eligibility based upon the following criteria: Healthy male nonsmokers, 20–79 years of age, normal serum PSA based upon the age- and race-specific cutoffs defined in the 2009 AUA Clinical Guidelines (15), no history or evidence of diabetes, PC, liver or kidney disease, and not taking >50 µg/day selenium as a dietary supplement.

Eligible subjects were randomly assigned with equal probability using a computer-generated list prepared by the biostatistician to one of 4 treatment groups: Placebo (non-selenized-yeast), low dose SY (SY{sub}200, 200 µg selenium/d), high dose SY (SY{sub}285, 285 µg selenium/d), and SeMet (200 µg selenium/d) (Figure 1) by the investigational pharmacists. The randomization status was blinded to all others in the study except to the Penn State University investigational pharmacists until a patient had finished the study and until near the end of the study upon IRB approval. The dose and form of selenium in the SY{sub}200 and SeMet groups were selected to match those used in the NPC (8) and SELECT (10) studies, respectively. The dose of selenium in the SY{sub}285 group was selected so that the levels of SeMet would be equivalent to that in the SeMet group, based upon a SeMet concentration in SY of 70% (16). Selenium-enriched yeast (SelenoExcel{superscript}®) was provided by Cypress Systems, Inc. (Madera, CA); SeMet was purchased from Sabinsa Corporation (Piscataway, NJ) and packaged by Cypress Systems, Inc. Placebo, SeMet, and SY capsules were identical in appearance. At baseline, a structured questionnaire was administered to each subject to collect information on demographics, medical history, medications, dietary supplements, alcohol consumption and cigarette smoking history. Study participants were remunerated with $35 per visit.

After randomization participants were provided their respective capsules along with instructions for usage. After 3, 6, and 9 months participants returned to the clinic to receive new and return unused capsules. At 9 months, all subjects were switched to placebo for the final 3 months of the trial (washout period). At baseline and after 3, 6, 9 and 12 months, biological samples including blood and urine were collected and processed as described below.

Outcome measures included plasma total selenium at baseline, and 3, 9 and 12 months; blood free and protein bound GSH and urinary 8-OHdG and 8-iso-PGF{sub}2α at baseline and 9 and 12 months. Secondary outcomes included blood glucose and serum PSA at baseline, and 3, 6, 9 and 12 months as well as selenium speciation in plasma at baseline and 9 months.

**Collection and processing of biological samples**

Fasting venous blood samples were collected between 9:00 am and 3:30 pm into either plain or EDTA-containing tubes and immediately placed on ice. An aliquot of whole blood was removed from the EDTA tube for analysis of glutathione. Remaining samples were centrifuged at 4°C and resulting plasma or serum was aliquoted and immediately frozen at −80°C. Packed red cells were washed 3× in saline and stored at −80°C. Urine was placed immediately on ice, aliquoted and frozen at −80°C.
**Analysis of blood and urine markers**

*Plasma total selenium levels* were determined by atomic absorption spectrophotometry as described previously (17).

**Selenium speciation in plasma**—Plasma SeMet, methylselenocysteine (MSC), selenate and selenite were analyzed by Ion Chromatography Inductively Coupled Plasma Collision Reaction Cell Mass Spectrometry by Applied Speciation and Consulting, LLC (Bothell, WA).

**Glutathione and glutathionylated proteins**—Total glutathione (GSH and glutathione disulfide) and protein bound GSH was analyzed as described previously (18, 19). Hemoglobin was determined by spectrophotometrically using Drabkin’s reagent (20).

**8-Iso-prostaglandin-F\(_2\)α (8-iso-PGF\(_2\)α) and 8-hydroxy-2′-deoxyguanosine (8-OHdG)** in urine were analyzed by ELISA Cayman Biochemical (Cat. Nos. 516351 and 589320; Ann Arbor, MI, USA). Creatinine was determined by reaction with picrate as described previously (21).

**PSA and glucose**—Serum PSA was determined using the ADVIA Centaur XP Immunoassay System (Siemens Healthcare Diagnostics Inc., Deerfield, IL). Blood glucose levels were determined using an Ortho Vitros 5600 analyzer.

**Statistical analysis**

Comparison of plasma selenium levels at each time point between groups was performed by Analysis of Variance with adjustment of multiple comparisons using Tukey’s method. Plasma levels of selenium metabolites were compared between baseline and 9 months using the Welch Two Sample t-test (22). Changes in serum PSA, plasma glucose and blood GSH and protein bound GSH and urine 8-isoprostane and 8-OHdG levels from baseline at different time points were compared between the 4 treatment groups using Analysis of Variance. Changes from baseline for urinary 8-OHdG and 8-isoprostane at 9 months were compared for each treatment group by baseline plasma selenium level tertile using Analysis of Variance.

**Results**

**Study subject characteristics**

The clinical phase of the study was conducted from May, 2008 until June, 2013. A total of 130 subjects were randomized, of which 69 were included in the analysis (Figure 1). Of the 61 randomized subjects not included in the analysis, 25 discontinued intervention prior to completion, 8 were lost to follow-up, 25 had not completed the protocol as of June 1, 2013 and 3 were excluded from analysis due to lack of compliance (Figure 1). The characteristics of these study participants are summarized in Table 1. All subjects were non-smokers (nonsmoker for at least for 1 yr prior to entry into the study) and had no history of chronic disease or high dose antioxidant supplement usage. Subjects ranged in age from 23 to 78 years of age (mean ± SD = 51.1 ± 14.0 years). No significant differences were observed.
between subjects by age, weight or BMI between treatment groups. A total of 70% of subjects fell within the normal to overweight categories with 30% being obese (BMI > 30).

**Compliance and Adverse Effects**

A total 5 of subjects (2 in the placebo group, 2 in the SY\textsubscript{285} group and 1 in the SeMet group) were removed from protocol due to adverse events: High PSA (1, SY\textsubscript{285} group), PC (1 in SY\textsubscript{285} group and 1 in SeMet group), and high blood glucose levels (2 in Placebo group). Among the participants that completed the study, no serious adverse effects were reported regardless of arm. All potential adverse events reported were minor (e.g., headaches and colds) and none were attributed to protocol treatment. Compliance was consistently high when assessed by either pill count or patient diary entries. Percent compliance was 97.0 ± 4.7 (mean ± SD) in all subjects and did not differ between treatment arms (placebo, 95.9 ± 5.4; SY\textsubscript{200}, 97.8 ± 3.9; SY\textsubscript{285}, 96.1 ± 6.3; SeMet, 98.0 ± 3.1).

**Plasma selenium levels**

Plasma selenium levels were not different between groups at baseline (Figure 2) and were consistent with those reported in the SELECT trial (10). In all selenium-treated groups, there was a progressive increase in levels from 3 to 9 months followed a return to baseline after the 3-month washout. In the SY groups, the increases were dose dependent reaching a maximum increase above baseline of 86% in the SY\textsubscript{285} group and 54% in the SY\textsubscript{200} group. When comparing the change from baseline between the four groups for months 3, 9 and 12 separately, significant differences were observed between all 3 selenium groups and placebo (P<0.0001) and between both the SeMet and SY\textsubscript{285} groups and the SY\textsubscript{200} group, respectively (P<0.05). At 12 months, statistical differences were observed between both SeMet and SY\textsubscript{285} groups and placebo only (P<0.001).

**Plasma Selenium Speciation**

Selenium speciation analyses were conducted in plasma samples collected at baseline and after 9 months from a subset of subjects randomly selected from the SeMet and high-dose SY groups (n=5 per group) (Table 2). These groups were selected based upon the equivalent doses of SeMet. Selenite was detected in all samples, whereas, selenate and MSC levels were below the limits of detection. SeMet was not detected in baseline samples but was detected in all 9-month samples. After 9 months of supplementation, significant increases from baseline were observed for SeMet and selenite in both groups (P<0.001). Overall, increases in SeMet and selenite could only account for 0.8% and 5.1%, respectively, of the increases observed for selenium at 9 months. Increases in SeMet levels from undetectable at baseline to 0.74 ng/ml in the SeMet group and 0.55 ng/ml in the SY\textsubscript{285} group were not significantly different between groups. Likewise, increases in selenite levels of 166% in the SeMet group and 88% in the SY\textsubscript{285} group were not significantly different between groups.

**Biomarkers of oxidative stress**

Levels of 8-iso-PGF\textsubscript{2α} in urine were measured as a biomarker of lipid peroxidation (Figure 3A, upper panel). A dose responsive decrease was observed for the SY groups reaching a maximum decrease of 28% occurring in the SY\textsubscript{285} group after 9 months (P<0.05). Levels
returned to baseline after the 3 month washout. No significant changes were observed in the SeMet, SY<sub>200</sub> or placebo groups.

8-OHdG levels in urine were measured as a biomarker of oxidative damage to DNA (Figure 3A, lower panel). A dose responsive decrease was observed for the SY groups reaching a maximum decrease of 34% in the SY<sub>285</sub> group after 9 months (P<0.05). Levels returned to baseline after the 3 month washout. No significant changes were observed in the SeMet, SY<sub>200</sub> and placebo groups.

The impact of baseline selenium levels on biomarkers of oxidative stress

Mean changes in urinary 8-OHdG and 8-iso-PGF<sub>2α</sub> levels at 9 months were compared between baseline selenium tertile groups (Supplementary Table 1). Significant associations were observed in the SY<sub>285</sub> group only (Figure 3B). In the lowest tertile of baseline selenium (mean ± SD: 115 ± 10.3 mg/ml) levels of both biomarkers were significantly reduced. In the middle tertile (136 ± 1.6 ng/ml), a significant reduction was observed for 8-iso-PGF<sub>2α</sub> only. No changes were observed in the highest tertile (152 ± 13.1 ng/ml). The largest decreases in both biomarkers were observed for the lowest tertile.

Glutathione and protein glutathionylation

Levels of free and protein bound GSH were measured in whole blood at baseline and after 9 and 12 months (Table 3). There were no changes in either free or bound levels of GSH were observed in any of the 4 study groups.

Glucose and PSA

At baseline, all subjects had normal fasting blood glucose and serum PSA levels; no differences were observed for either glucose or PSA between groups (Table 3). In all groups, glucose and PSA levels were each unchanged during the course of the study.

Discussion

This is the first clinical trial aimed at comparing the effect of two different forms of selenium on biomarkers that are likely to influence PC risk in men. In a randomized double-blind, placebo-controlled trial in healthy men, we have observed that SY but not SeMet (the formulation used in the largest PC chemoprevention study (10) ever conducted) was effective at reducing the levels of oxidative stress as assessed by reductions in biomarkers of lipid peroxidation (8-iso-PGF<sub>2α</sub>) and oxidative damage to DNA (8-OHdG), despite inducing similar increases in plasma selenium levels. Although the mechanisms of chemoprevention by selenium remain unclear, enhanced protection against oxidative stress is thought to be involved (23–27). Selenium supplementation in the form of sodium selenite was previously shown to decrease biomarkers of oxidative stress (lymphocyte 8-oxodG and urinary excretion 8-oxoGua) in individuals carrying the BRCA1 mutation (28). This difference in effectiveness of SY vs. SeMet in protecting against oxidative stress may, in part, help explain the results of previous trials which demonstrated the effectiveness of SY (NPC) (8), but not SeMet (SELECT) (10), at reducing the risk for PC. In fact SY was the only form associated with a pattern of decreased DNA damage in rodents (14).
In the present study we observed that both SeMet and SY increased plasma selenium levels and, for SY, the increases were dose-dependent. The increases observed for the SY\textsubscript{285} group were comparable to the SeMet group, despite the total dose of selenium being 85 µg/day greater in the SY\textsubscript{285} group. Since the dose of SeMet were the same in both the SeMet and SY\textsubscript{285} supplements, these results suggest that SeMet is the form of selenium in yeast that is responsible for enhancing plasma total selenium levels. These results are consistent with previous findings which indicate that SeMet-containing proteins (where SeMet is incorporated non-specifically in place of Met), the most abundant source of selenium in plasma (29–32), are the major source for selenium variation in plasma and that selenoproteins or other forms of selenium (eg. MSC) account for very little of the variation of selenium in plasma (33–35). While we were not able to measure the impact of SY or SeMet on prostate selenium status, in a recent clinical study, SY supplementation was associated with a dose-dependent increase in total selenium levels in prostatic tissues that appeared to be greater than that previously observed with SeMet (36). Collectively, these results indicate that the selenium disposition in plasma may differ from that in the target organ (prostate).

Our current finding that the impact of SY on oxidative stress biomarkers was greatest in men with low baseline plasma selenium levels are consistent with a recent report of SY supplementation in healthy men (37) and provides further support for the importance of baseline selenium in predicting the efficacy of supplementation. Epidemiological investigations have linked low selenium intake and levels with increased risk for PC (38–40) including the recent prospective study in the Netherlands, where a strong association between toenail selenium levels and advanced PC was observed in a population with relatively low levels selenium (6). In the NPC, the greatest protective effects were observed in individuals with low baseline selenium and no beneficial effects were observed in individuals with levels above 122 µg/L (9). This is consistent with the present trial where optimal beneficial effects on oxidative stress biomarkers were observed in individuals in the lowest tertile for baseline plasma selenium (≤27 µg/L). The higher baseline levels of selenium in SELECT participants compared to those in NPC may contribute to the lack of a protective effect of selenium observed in that trial. In a recent reevaluation of SELECT, no association of PC risk was observed with baseline toenail selenium levels (41). However, the baseline levels of toenail selenium in this trial were on average >60% higher than in those in the Netherlands trial such that >80% of the subjects in the Netherlands trial would fall within the lowest quintile of SELECT. Considering the baseline selenium levels observed in these trials, Rayman et al. pointed out that the results of NPC are consistent with those of SELECT (42). The lack of a protective effect of SY (200 µg/day) on PC in high risk men in the recently completed Negative Biopsy Trial may be due to the late stage of disease when intervention was started but may also reflect the relatively higher baseline levels of plasma selenium observed in this trial compared to NPC (43).

While SeMet does not appear to be the active chemopreventive agent in SY, it is currently not known which agents may be responsible for this activity. Interest in MSC as one such agent stems from preclinical studies which show that it is more effective than SeMet at impacting PC related pathways in preclinical models (5). We recently identified a number of proteins that are differentially expressed as a result of selenium enrichment of yeast (44),
including the selenium-containing protein elongation Factor 2 (45) and others have identified up to 27 selenium containing compounds in SY (46) which may also be playing a role.

A limitation of this study was the relatively small sample size, especially among men with low selenium at baseline, although adequately powered as a randomized phase II study that can support future larger studies. Also, outcome measures did not include PC development or biomarkers specific for PC risk. Strengths of the study include the randomized double-blind, placebo-controlled design in healthy men and the use of selenium supplements at doses which mimic previously conducted disease outcome trials and also allow for comparison between different selenium forms.

Overall, our study highlights the differences between the effects of SY and SeMet in preventing oxidative stress and support the continued development of SY but not SeMet as a chemopreventive agent. Because the effects of SY appear to be based on baseline levels of plasma selenium, such a chemopreventive approach may be best served if applied to men with low selenium levels as suggested in a recent study (6).

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

The authors acknowledge the assistance of the staff of the Community Sciences and Health Outcomes Core and Clinical Trials Office of the Penn State Hershey Cancer Institute in this research. We also thank the nursing staff in the Clinical Research Center, Christopher Hamilton in the Core Endocrine Laboratory, Cheryl Reitzel in the Clinical Trials Office for her assistance with Oncore, and Heather Heisey and Alyse Fazzi in the Investigational Drug Service of the Department of Pharmacy, Penn State University College of Medicine. We also acknowledge the contribution of the staff of the Office of Human Research Services at the Rutgers Cancer Institute of New Jersey and the nursing staff at Clinical Research Center, Penn State University, University Park, PA. We thank Dr. Telih Boyiri for his assistance with all regulatory aspects of this trial.

Financial Support: This work was supported a National Cancer Institute grant (R01CA127729) (PI: KE-B). Selenium speciation analyses were supported by Cypress Systems, Inc. (Madera, CA). Additional clinical support was provided by the Penn State Hershey Cancer Institute through its Clinical Trials Office and Community Sciences and Health Outcomes Core and the Office of Human Research Services at the Cancer Institute of New Jersey.

Abbreviations

<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>SY</td>
<td>selenium-enriched yeast</td>
</tr>
<tr>
<td>SeMet</td>
<td>selenomethionine</td>
</tr>
<tr>
<td>GSH</td>
<td>glutathione</td>
</tr>
<tr>
<td>8-OHdG</td>
<td>8-hydroxy-2’-deoxyguanosine</td>
</tr>
<tr>
<td>8-iso-PGF2α</td>
<td>8-iso-prostaglandin-F2α</td>
</tr>
<tr>
<td>PC</td>
<td>prostate cancer</td>
</tr>
<tr>
<td>NPC</td>
<td>Nutritional Prevention of Cancer</td>
</tr>
</tbody>
</table>

Cancer Prev Res (Phila). Author manuscript; available in PMC 2015 August 01.
SELECT Selenium and Vitamin E Cancer Prevention Trial
SY200 200 µg selenium/d
SY285 285 µg selenium/d
MSC methylselenocysteine

References


Cancer Prev Res (Phila). Author manuscript; available in PMC 2015 August 01.
Figure 1.
Subject flowchart summarized according to the Consolidated Standards of Reporting Trials.
Figure 2.
Effect of supplementation with SeMet or SY on plasma selenium levels in adult men. Subjects were randomized to placebo (n=18), 200 µg/d SY (n=16), 285 µg/d SY (n=15) or 200 µg/d SeMet (n=20). Supplementation continued for 9 months followed by a 3 month washout. Plasma total selenium levels were assessed by atomic absorption spectrophotometry. Symbols and bars represent mean and standard error values, respectively.

* significantly different from placebo, P<0.0001
† significantly different from low dose SY group, P<0.005
‡ significantly different from low dose SY group, P<0.05
Figure 3.
Effect of supplementation with SeMet or SY on urinary biomarkers of oxidative stress in healthy men. Subjects were randomized to placebo (n=18), 200 µg/d SY (n=16), 285 µg/d SY (n=15) or 200 µg/d SeMet (n=20). Urinary levels of 8-iso-PGF$_{2\alpha}$ and 8-OHdG were determined by ELISA and corrected for urinary dilution by dividing by urinary creatinine levels, and plasma selenium, determined by atomic absorption spectrophotometry. A. Urinary levels of 8-iso-PGF$_{2\alpha}$ (upper panel) and 8-OHdG (lower panel) by treatment arm. Bars represent mean change in biomarker values from baseline after 9 and 12 months with associated standard error values. Baseline values for 8-iso-PGF$_{2\alpha}$ were 199 ± 96, 180 ± 87, 217 ± 117, and 206 ± 114 mg/mg creatinine for placebo, SY$_{200}$, SY$_{285}$ and SeMet groups, respectively. Baseline values for 8-OHdG were 66.8 ± 22, 72.7 ± 43, 90.1 ± 38, and 70.1 ± 38 mg/mg creatinine for placebo, SY$_{200}$, SY$_{285}$ and SeMet groups, respectively. B. Association of reductions in oxidative stress biomarker levels by SY with baseline selenium levels in plasma. Urinary levels of 8-iso-PGF$_{2\alpha}$ and 8-OHdG for subjects in the 285 µg/d SY group were examined by baseline plasma selenium tertile. Mean baseline plasma selenium levels by tertile are provided in the top panel. Mean changes in 8-OHdG (middle panel) and 8-iso-PGF$_{2\alpha}$ (bottom panel) from baseline at 9 months are reported by baseline selenium tertile. Bars represent mean and standard error values. Tertile baseline plasma cutpoints (mg/ml) are 127 and 138.
Table 1

Study Subject Characteristics at Baseline

<table>
<thead>
<tr>
<th>Group</th>
<th>N</th>
<th>Age (yr)</th>
<th>Race/Ethnicity n (%)</th>
<th>BMI (kg/m²)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Placebo (plain yeast)</td>
<td>18</td>
<td>48.1 ± 14.6 (22–70)</td>
<td>White: 17 (94%)</td>
<td>30.0 ± 4.79 (22.0–38.3)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Asian: 0 (0%)</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Black: 1 (6%)</td>
<td></td>
</tr>
<tr>
<td>SY (200 µg/d)</td>
<td>16</td>
<td>50.7 ± 16.2 (23–78)</td>
<td>White: 15 (94%)</td>
<td>28.0 ± 3.20 (22.4–34.9)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Asian: 0 (0%)</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Black: 1 (6%)</td>
<td></td>
</tr>
<tr>
<td>SY (285 µg/d)</td>
<td>15</td>
<td>51.3 ± 12.0 (25–72)</td>
<td>White: 13 (87%)</td>
<td>27.8 ± 3.14 (23.7–34.2)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Asian: 0 (0%)</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Black: 1 (7%)</td>
<td></td>
</tr>
<tr>
<td>SeMet (200 µg/d)</td>
<td>20</td>
<td>54.0 ± 13.4 (30–75)</td>
<td>White: 17 (85%)</td>
<td>28.5 ± 3.79 (23.0–36.4)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Asian: 2 (10%)</td>
<td></td>
</tr>
</tbody>
</table>

*Values are mean ± SD (range)
### Table 2

**Plasma Selenium Speciation**

<table>
<thead>
<tr>
<th>Analyte‡</th>
<th>Group</th>
<th>Baseline</th>
<th>9 Months</th>
<th>Change from baseline</th>
</tr>
</thead>
<tbody>
<tr>
<td>Selenium</td>
<td>SeMet</td>
<td>139 ± 7.0</td>
<td>256 ± 14.9†</td>
<td>117 ± 13.2</td>
</tr>
<tr>
<td></td>
<td>SY285</td>
<td>129 ± 8.7</td>
<td>256 ± 26.9†</td>
<td>128 ± 30.2</td>
</tr>
<tr>
<td>SeMet</td>
<td>SeMet</td>
<td>&lt;0.12</td>
<td>0.736 ± 0.16†</td>
<td>0.616 ± 0.16</td>
</tr>
<tr>
<td></td>
<td>SY285</td>
<td>&lt;0.12</td>
<td>0.550 ± 0.07†</td>
<td>0.430 ± 0.07</td>
</tr>
<tr>
<td>Selenite</td>
<td>SeMet</td>
<td>2.97 ± 0.22</td>
<td>7.90 ± 0.37†</td>
<td>4.93 ± 0.42</td>
</tr>
<tr>
<td></td>
<td>SY285</td>
<td>4.47 ± 0.49</td>
<td>8.43 ± 0.32†</td>
<td>3.96 ± 0.24</td>
</tr>
</tbody>
</table>

*Values are mean ± SE (n=5).

†Significantly different from baseline (P<0.001).

‡Selenate and methylselenocysteine concentrations were below detection limits (0.12 ng/ml for selenate and 0.16 ng/ml for methylselenocysteine) in all samples.
Table 3

Effect of selenium-enriched yeast or SeMet on serum PSA and blood glucose and free and bound GSH in healthy men.

<table>
<thead>
<tr>
<th>Group</th>
<th>Baseline</th>
<th>3 mo</th>
<th>6 mo</th>
<th>9 mo</th>
<th>12 mo</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glucose (mg/dl)*</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Placebo</td>
<td>84.7 ± 2.29</td>
<td>1.28 ± 2.01</td>
<td>3.06 ± 2.32</td>
<td>3.72 ± 2.76</td>
<td>4.28 ± 1.79</td>
</tr>
<tr>
<td>SY (200 µg/day)</td>
<td>91.6 ± 1.65</td>
<td>0.12 ± 2.08</td>
<td>2.69 ± 2.62</td>
<td>−1.44 ± 2.75</td>
<td>−1.06 ± 2.70</td>
</tr>
<tr>
<td>SY (285 µg/day)</td>
<td>82.7 ± 2.45</td>
<td>0.40 ± 1.89</td>
<td>−3.67 ± 2.57</td>
<td>1.60 ± 5.31</td>
<td>1.00 ± 3.09</td>
</tr>
<tr>
<td>SeMet (200 µg/day)</td>
<td>91.0 ± 2.07</td>
<td>−0.50 ± 0.96</td>
<td>−1.45 ± 1.03</td>
<td>−1.75 ± 2.46</td>
<td>−2.20 ± 2.49</td>
</tr>
<tr>
<td>PSA (ng/ml)*</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Placebo</td>
<td>1.03 ± 0.12</td>
<td>0.07 ± 0.03</td>
<td>0.32 ± 0.22</td>
<td>0.37 ± 0.26</td>
<td>0.05 ± 0.06</td>
</tr>
<tr>
<td>SY (200 µg/day)</td>
<td>1.40 ± 0.29</td>
<td>0.14 ± 0.09</td>
<td>0.22 ± 0.13</td>
<td>0.34 ± 0.30</td>
<td>0.74 ± 0.50</td>
</tr>
<tr>
<td>SY (285 µg/day)</td>
<td>0.59 ± 0.06</td>
<td>0.05 ± 0.03</td>
<td>0.09 ± 0.04</td>
<td>0.08 ± 0.04</td>
<td>0.06 ± 0.05</td>
</tr>
<tr>
<td>SeMet (200 µg/day)</td>
<td>1.14 ± 0.16</td>
<td>−0.08 ± 9.09</td>
<td>0.06 ± 0.07</td>
<td>0.02 ± 0.08</td>
<td>0.02 ± 0.07</td>
</tr>
<tr>
<td>GSH (µmol/ml)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Placebo</td>
<td>0.94 ± 0.08</td>
<td>ND</td>
<td>ND</td>
<td>−0.12 ± 0.08</td>
<td>−0.09 ± 0.09</td>
</tr>
<tr>
<td>SY (200 µg/day)</td>
<td>0.90 ± 0.07</td>
<td>ND</td>
<td>ND</td>
<td>−0.05 ± 0.08</td>
<td>−0.01 ± 0.12</td>
</tr>
<tr>
<td>SY (285 µg/day)</td>
<td>0.94 ± 0.09</td>
<td>ND</td>
<td>ND</td>
<td>−0.02 ± 0.09</td>
<td>−0.10 ± 0.07</td>
</tr>
<tr>
<td>SeMet (200 µg/day)</td>
<td>0.99 ± 0.07</td>
<td>ND</td>
<td>ND</td>
<td>−0.22 ± 0.07f</td>
<td>−0.11 ± 0.08</td>
</tr>
<tr>
<td>Protein bound GSH (% of total GSH)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Placebo</td>
<td>17.7 ± 1.85</td>
<td>ND</td>
<td>ND</td>
<td>−1.37 ± 2.40</td>
<td>−1.14 ± 2.31</td>
</tr>
<tr>
<td>SY (200 µg/day)</td>
<td>16.5 ± 1.64</td>
<td>ND</td>
<td>ND</td>
<td>−0.39 ± 1.97</td>
<td>0.54 ± 2.43</td>
</tr>
<tr>
<td>SY (285 µg/day)</td>
<td>15.7 ± 1.33</td>
<td>ND</td>
<td>ND</td>
<td>1.86 ± 2.57</td>
<td>4.17 ± 2.09</td>
</tr>
<tr>
<td>SeMet (200 µg/day)</td>
<td>16.8 ± 1.61</td>
<td>ND</td>
<td>ND</td>
<td>1.25 ± 1.79</td>
<td>−0.51 ± 2.34</td>
</tr>
</tbody>
</table>

Subjects were randomized to placebo (n=18), 200 µg/d SY (n=16), 285 µg/d SY (n=15) or 200 µg/d SeMet (n=20). Values are mean ± SE.

* Eligibility criteria: Glucose <106 mg/dl; normal PSA based on age at enrollment ≤4.0 ng/ml

† Statistically significant from baseline (P<0.05).

ND: not determined